

NMR study of the calmodulin/Munc13-1 complex reveals multiple simultaneous binding modes

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The priming process is an essential and rate-limiting step for the post-docking/pre-fusion maturation of vesicles in regulated exocytosis [1]. The interactions between Munc13 and calmodulin (CaM) regulate the synaptic vesicle priming and synaptic efficacy in response to the residual Ca²⁺ signal, thereby shaping the short-term plasticity characteristics during periods of synaptic activity [2]. Moreover, it has been recently shown that the Munc13s C1 domain lowers the energy barrier for synaptic vesicle fusion [3].

Munc13-1 features a CaM binding site (Munc13-1⁴⁵⁸⁻⁴⁹²) located N-terminal to its C1 domain (Munc13-1⁵⁶⁷⁻⁶³¹), which binds CaM in a Ca²⁺-dependent manner. The NMR solution structure of the CaM/Munc13-1⁴⁵⁸⁻⁴⁹² complex, presented here, features a novel (1-5-8-25-26) CaM binding motif, where the Munc13-1 residues W464, F468 and V471 anchor the protein to the C-terminal CaM domain, while L488 and W489 are attached to the N-terminal CaM domain. This structure features a flexible arrangement between both CaM domains, as no inter-domain NOEs were observed.

Additionally, we demonstrate the presence of an interaction between the C1 domain of Munc13-1 and CaM, which occurs in the NMR fast time scale. Chemical shifts perturbations (CSPs) and paramagnetic pseudo-contact shifts (PCSs) clearly identify the N-terminal CaM domain as the one binding the Munc13-1 C1 domain, while no interactions between Munc13-1⁴⁵⁸⁻⁴⁹² and the C1 domain were observed. Altogether, these results indicate that CaM binds simultaneously to the CaM-binding site and the C1 domain of Munc13-1. Thereby, the C-terminal CaM domain plays a leading role in the complex stabilization, through a high affinity interaction with the CaM-binding site of Munc13-1, while the N-terminal CaM domain seems to have a regulatory function by interacting simultaneously with the L488-W489 segment and the C1 domain.

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